



# Goat anti-Chicken IgY (H+L) Secondary Antibody, Alexa Fluor™ 594

<b>Product Details</b>	
Size	1 mg
Species Reactivity	Chicken
Host/Isotype	Goat / IgG
Class	Polyclonal
Туре	Secondary Antibody
Conjugate	Alexa Fluor™ 594
Excitation/Emission Max	590/618 nm
Immunogen	Gamma Immunoglobins Heavy and Light chains
Form	Liquid
Concentration	2 mg/mL
Purification	purified
Storage buffer	PBS, pH 7.5
Contains	5mM sodium azide
Storage conditions	4° C, store in dark
RRID	AB_2534099

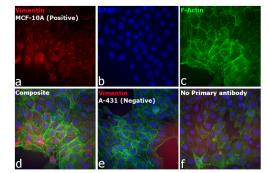
Applications	Tested Dilution	Publications
Immunohistochemistry (IHC)	1-10 μg/mL	0 Publication
Immunohistochemistry (Paraffin) (IHC (P))	-	0 Publication
Immunohistochemistry (Frozen) (IHC (F))	-	0 Publication
Immunohistochemistry - Free Floating (IHC (Free))	-	0 Publication
Immunocytochemistry (ICC/IF)	1-10 μg/mL	0 Publication
Flow Cytometry (Flow)	1-10 μg/mL	-
Miscellaneous PubMed (Misc)	-	0 Publication

#### **Product Specific Information**

Alexa Fluor dyes are among the most trusted fluorescent dyes available today. Invitrogen™ Alexa Fluor 594 dye is a bright, red-fluorescent dye with excitation ideally suited to the 594 nm laser line. For stable signal generation in imaging and flow cytometry, Alexa Fluor 594 dye is pH-insensitive over a wide molar range. Probes with high fluorescence quantum yield and high photostability allow detection of low-abundance biological structures with great sensitivity. Alexa Fluor 594 dye molecules can be attached to proteins at high molar ratios without significant self-quenching, enabling brighter conjugates and more sensitive detection. The degree of labeling for each conjugate is typically 2-8 fluorophore molecules per IgG molecule; the exact degree of labeling is indicated on the certificate of analysis for each product lot.

Using conjugate solutions: Centrifuge the protein conjugate solution briefly in a microcentrifuge before use; add only the supernatant to the experiment. This step will help eliminate any protein aggregates that may have formed during storage, thereby reducing nonspecific background staining. Because staining protocols vary with application, the appropriate dilution of antibody should be determined empirically. For the fluorophore-labeled antibodies a final concentration of 1-10 µg/mL should be satisfactory for most immunohistochemistry and flow cytometry applications.

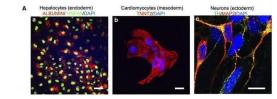
## Product Images For Goat anti-Chicken IgY (H+L) Secondary Antibody, Alexa Fluor™ 594



Chicken IgY (H+L) Secondary Antibody (A-11042) in ICC/IF Immunofluorescence analysis of Goat anti-Chicken IgY (H+L) Secondary Antibody, Alexa Fluor™ 594, (Product # A-11042) was performed using MCF 10A (positive model) and A-431 (negative model) cells stained with Vimentin Polyclonal antibody (Product # PA1-10003). The cells were fixed with 4% paraformaldehyde for 10 minutes, permeabilized with 0.1% Triton™ X-100 for 10 minutes, blocked with 2% BSA for 1 hour and labeled with 1:800 dilution of primary antibody at 4 degree celsius. Goat anti-Chicken IgY (H+L) Secondary Antibody, Alexa Fluor™ 594, (Product # A-11042, 1:2000 dilution) in 0.1% BSA in PBS for 45 minutes at room temperature, was used for detection of Vimentin in the cytoskeleton (Panel a: Red). Nuclei (Panel b: blue) were stained with Hoechst33342 (Product # H1399). F-actin was stained with Alexa Fluor® 488 Phalloidin (Product # A12379, 1:500) (Panel c: green). Panel d represents the composite image. The specificity of the secondary antibody was proved by the absence of signal in A-431 (negative model for Vimentin) due to no primary antibody binding (Panel e). Non-specific staining was not observed with secondary antibody alone (panel f). The images were captured at 40X magnification in CellInsight CX7 LZR High-Content Screening (HCS) Platform (Product # CX7A1110LZR) and externally deconvoluted (D.Sage et al./Methods 115 (2017) 28-41).

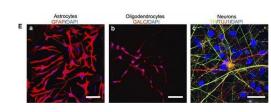
#### Chicken IgY (H+L) Secondary Antibody (A-11042) in ICC/IF

POLG iPSCs manifested a partial phenotype presenting with energy depletionARepresentative confocal images of iPSC lineage-specific differentiation into germ layers of endoderm-derived hepatocytes with positive expression of ALBUMIN (red) and HNF4A (green) (a) (scale bar, 100m), mesodermal-derived cardiomyocytes with positive expression of TNNT2 (red) (b) (scale bar, 100m), and ectodermal-derived dopaminergic neurons with positive expression of TH (green) and MAP2 (red) (c) (scale bar, 10m). Nuclei are stained with DAPI (blue). BConfocal images of mitochondrial morphology for iPSC lines with co-staining of MTG (upper panel) and TMRE (lower panel) (scale bars, 25m). Nuclei are stained with DAPI (blue). C-EFlow cytometric analysis of iPSCs generated from Detroit 551, WS5A, and CP2A fibroblasts for mitochondrial volume (MTG) (C, n=6, technical replicates per clone for control and CP2A; n=5, technical replicates per clone for WS5A), total MMP (TMRE) (D. n=6, technical replicates per clone for control and CP2A; n=5, technical replicates per clone for WS5A) and specific MMP (E, n=6, technical replicates per clone for control and CP2A; n=5, technical replicates per clone for WS5A) calculated by dividing median fluorescence intensity (MFI) for total TMRE expression by MTG. FIntracellular ATP production in iPSCs generated from Detroit 551, WS5A, and CP2A fibroblasts (n=3, technical replicates per clone for control ... Image collected and cropped by CiteAb from the following publication (https://pubmed. ncbi.nlm.nih.gov/32840960), licensed under a CC BY license.



## Chicken IgY (H+L) Secondary Antibody (A-11042) in ICC/IF

NSCs retained the original genotype and sequenceAQuantification of gene expression for NSC markers PAX6, NESTIN, and SOX2 for all NSCs from RTqPCR analysis. The expression of the neural stem cell markers is assessed with fold change using the comparative Ct method by normalizing NSCs to iPSCs (n=3, technical replicates per ESC line or iPSC clone).B, CQuantification of protein expression level for PAX6 (B, n=6, technical replicates per line for ESCs; n=7, technical replicates per clone for control; n=8, technical replicates per clone for WS5A; n=9, technical replicates per clone for CP2A) and NESTIN (C, n=6, technical replicates per line for ESCs; n=7, technical replicates per clone for control; n=8, technical replicates per clone for WS5A; n=9, technical replicates per clone for CP2A) for iPSC-derived NSCs using flow cytometry. DSequencing chromatogram showing the homozygous c.2243G>C variation in POLG in WS5A iPSC-derived NSCs and the heterozygous c.1399G>A and c.2243G>C variation in POLG in CP2A iPSC-derived NSCs. ERepresentative confocal images showing glial and neuronal lineages derived from NSCs. (a) Immunostaining of NSCderived astrocytes with GFAP (red) staining (scale bar, 50m). (b) Immunostaining of oligodendrocytes showing GALC (red)-positive labeling (scale bar, 50m). (c) Dopaminergic neurons showing TH (green) and TUJ1 (red)-positive staining (scale bar, 25m). Nuclei are stained with DA... Image collected and cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov /32840960), licensed under a CC BY license.



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#### □ 254 References

Fos expression in the periaqueductal gray, but not the ventromedial hypothalamus, is correlated with psychosocial stress-induced cocaine-seeking behavior in rats bioRxiv (2025)

Keratinocyte-derived extracellular vesicles in painful diabetic neuropathy. Neurobiol Pain (2025)

Transgenic A53T mice have astrocytic -synuclein aggregates in dopamine and striatal regions bioRxiv (2025)

The C-terminus of the TlpD cytoplasmic chemoreceptor promotes polar localization and function bioRxiv (2024)

Systemic Inflammation Decreases Initial Brain Injury but Attenuates Neurite Extension and Synapse Formation during the Repair of Injured Brains. Exp Neurobiol (2024)

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